0960-894X/96 \$15.00 + 0.00

PII: S0960-894X(96)00243-0

AZONIA DERIVATIVES OF THE γ-CARBOLINE SYSTEM. A NEW CLASS OF DNA INTERCALATORS

Andrés Molina, Juan J. Vaquero, José L. García-Navio, Julio Alvarez-Builla*, María M. Rodrigo, Obis Castaño, José L. de Andres

Departamentos de ^aQuímica Orgánica y ^bQuímica-Física, Universidad de Alcalá, 28871-Alcalá de Henares, Madrid. Spain. FAX 341 885 4686. E-mail JALVAREZ@QUIMOR.ALCALA.ES.

^cInstitut de Quimica Computacional, Universitat de Girona. C/Albareda, 3-5. 17071-Girona, Spain.

Abstract: 1-Methyl-γ-carboline derivatives were transformed into the corresponding N-aminoazinium salts, which were condensed with 1,2-dicarbonyl compounds (Westphal reaction) to afford azonia derivatives with a bridgehead quaternary nitrogen atom. Some of them show DNA intercalating properties. Copyright © 1996 Elsevier Science Ltd

Since DNA is an important cellular receptor, most anticancer agents exert their effects trough binding to it. Intercalation is one of these modes of interaction, in which a molecule is inserted between two adjacent base pairs causing lengthening, stiffening and unwinding of the helix. Planar polyheteroaromatic cations are particularly well adapted for intercalation between nucleic acid base pairs. While the positive charge in the chromophore seems to be essential for increasing their DNA affinity, their orientation in the intercalative process is highly dependent on the electrostatic component of the stacking interaction. Favourable dipolar interactions between the chromophore and the base pairs contribute to explain the observed selectivity for CpG or TpA steps. For these reasons, alterations of the dipole moment of the chromophore is an important element to be considered when DNA bisintercalators are being designed.

Figure 1

The above considerations led us to explore the DNA binding properties of type 1 azonia derivatives, incorporating as chromophore a polyheteroaromatic cation, in which the positive charge is not introduced by N-alkylation of the azaheterocycle, such is the case of well known DNA intercalators, such as ethidium bromide and some antitumour compounds of the ellipticine group (e. g. Céliptium®), but by the presence of a quaternary bridgehead nitrogen. In this communication we wish to report our initial results on the transformation of the γ -carboline system into tetracyclic and heptacyclic azonia derivatives, along with the DNA binding properties and charge distribution of a representative compound.

The 1-methylcarbolines were prepared by thermal or microwave decomposition of the corresponding pyridylbenzotriazoles 2 (Graebe-Ullmann reaction)⁵ as previously described.⁶ These derivatives were easily transformed into the salts 3 by amination with (O-mesitylenesulfonyl)hydroxylamine (MSH) in CH₂Cl₂ at room

temperature. The reaction of 3 with various 1,2-dicarbonyl compounds (Westphal condensation)⁷ afforded several types of azonia derivatives (Scheme 1). Different reaction conditions were necessary for the condensation to be successful. Thus, to obtain the polycyclic cations 5 and 6 from the reaction of 3 with 1,2-acenaphthoquinone and 9,10-phenanthrenequinone, the condensation had to be carried out in sodium acetate/acetone whereas in the reaction of 3 with 2,3-butanedione or 3,4-hexanedione, 4a-d could only be obtained if triethylamine/MeOH/ acetone was employed. The derivative 4e required the use of dibutylamine/EtOH.

Scheme 1. i) 150-200° C or MW, 160 watts; ii) MSH / CH₂Cl₂; iii) R⁴COCOR⁴, Et₃N / EtOH-acetone or Bu₂NH / EtOH; iv) 1,2-Acenaphthoquinone, NaOAc / acetone; v) 9,10-Phenanthrenequinone, NaOAc / acetone

Several techniques⁸ were used to evaluate the DNA binding properties of azonia derivatives 4-6. Whereas the addition of compounds 5 and 6 to a sample of calf thymus DNA in TRIS-HCl buffer ([NaCl]=0.05 M, pH=7.5) induced hyperchromicity, probably due to dipole-dipole interactions, the cationic derivatives 4 gave hypochromic and bathochromic shifts in their UV spectra (see Table) accounted for by their binding to DNA. The unique binding mode with DNA was established by the appearance of isosbestic points for this series of compounds. From the spectral changes, nonlinear Scatchard binding isotherms were generated. The McGhee-Von Hippel¹⁰ treatment allowed the values of K (DNA affinity constant) and n (number of base-pairs occluded by each bound ligand molecule) to be determined. Intercalation was proved by measuring the viscosity of sonicated calf thymus DNA in the absence and presence of increasing concentrations of 4. The slope¹¹ (m) obtained by plotting the relative increase in contour length (L/L_0) against the ligand binding ratio (r) gave values within the range of well known intercalators such as ethidium bromide, ¹² used here as reference. In the Table are shown the values for each compound¹³ along with their cellular toxicity against colon carcinoma HT-29 cells. Although the number of analised compounds is small, the presence and position of alkyl groups appears to have only a small effect on K values. EC₅₀ values indicate that inhibitory activity is enhanced by alkyl substitution, with the tetrasubstituted derivative 4e being the most potent of this series whilst the monosubstituted derivatives 4a and 4c were significantly less potent.

To be used in the comparison with other DNA intercalators, the optimized geometries and atomic charges of the cation 4a and the betaine 7 were determined using *ab initio* theoretical techniques¹⁵ (HF/6-31G*// HF/3-21G) and are presented in Figure 2. Calculations show that both are planar structures, determined as minima by frequency calculations. As a result, the easily formed and highly stable compound 7 seems better represented by the resonance hybrid 7-8 (Scheme 2).

Table. DNA Binding Properties of Azonia Derivatives 4

Compd. No	λ_{free}	λ_{bound}	$\epsilon_{ ext{free}}$	$\epsilon_{ ext{bound}}$	$\lambda_{ ext{\tiny IP}}$	10 ⁵ K	n	m	EC ₅₀ [μM]
4a	354	364	7637	4845	388	1.10	4.75	1.16	>10
4b	358	368	7187	4461	388	2.39	2.84	0.97	2.0
4c	352	362	7451	5056	390	1.27	3.98	0.95	>10
4d	356	366	7036	4651	388	2.62	3.56	0.87	2.1
4e	368	384	5887	3547	398	1.45	3.00	0.77	0.5
EtBr						12.0	2.00ª	1.11	

K: affinity constant for DNA (M⁻¹). n: Number of base-pairs occluded by each bound ligand molecule. m: Helix extension slope measured by sonicated DNA viscosimetric lengthening; values within 0.09-0.13 error. λ_{free} and λ_{bound} : Wavelength of maximum absorption for free and DNA bound compounds. ϵ_{free} and ϵ_{bound} : Extinction coefficients for free and DNA bound compounds. λ_{IP} : Wavelength of isosbestic point. EC₅₀: Concentration of drug required to inhibit 50% of the cell growth after 72 h, of colon carcinoma HT-29. In vitro activity was measured as indicated in ref. 14 (doxorubicin, EC₅₀=2.5). EtBr: Ethidium bromide.

In summary, this work presents a series of new pyridazino[1',6':1,2]pyrido[4,3-b]indol-5-inium salts which shown both batho- and hypochromicity in their UV spectra in the presence of DNA and increased viscosity in sonicated DNA. Transformation into bis-salts is now under way in order to improve on the observed affinity constants in these new DNA intercalators.

Acknowledgements. Financial support from the Comisión Interministerial de Ciencia y Tecnología (CICYT, project SAF94-0280) and studentships (A.M.) from the Ministerio de Educación y Ciencia are gratefully acknowledged. We also thank the Institut de Química Computacional for the computing facilities.

^{*} Value according to neighbor exclusion model (ref. 16)

References and Notes

- Wilson, W. D.; Jones, R in *Intercalation Chemistry*, Whittingham, M. S.; Jacobson, A. J. Eds.; Academic Press, New York, 1981; Chapter 14.
- 2. Gallego, J.; Ortiz, A. R.; Gago, F. J. Med. Chem. 1993, 36, 1548.
- (a) Gallego, J.; Luque, F. J.; Orozco, M.; Burgos, C.; Alvarez-Builla, J.; Rodrigo, M. M.; Gago, F. J. Med. Chem. 1994, 37, 1602.
 (b) Gallego, J.; Luque, F. J.; Orozco, M.; Gago, F. J. Biomol. Str. & Dyn. 1994, 12, 111.
- 4. Pastor, J.; Siró, J.; García-Navío, J. L.; Vaquero, J. J.; Rodrigo, M. M.; Ballesteros, M.; Alvarez-Builla, J. Bioorg. Med. Chem. Lett. 1995, 5, 3043.
- 5. Graebe, C.; Ullman, F. Justus Liebigs Ann. Chem. 1896, 291, 16.
- For a preliminary communication: Molina, A.; Vaquero, J. J.; Garcia-Navio, J. L.; Alvarez-Builla, J. Tetrahedron Lett. 1993, 34, 2673.
- 7. For recent examples see: (a) Santiesteban, I.; Siro, J. G.; Vaquero, J. J.; García-Navio, J. L.; Alvarez-Builla, J.; Castaño, O.; Andrés, J. L. J. Org. Chem. 1995, 60, 5667. (b) Diaz, A.; Matia, M. P.; García-Navío, J. L.; Vaquero, J. J.; Alvarez-Builla, J. J. Org. Chem. 1994, 59, 8294.
- 8. For criteria on the mode of binding to DNA see: (a) Suh, D.; Chaires, J. B. *Bioor. Med Chem.* **1995**, *3*, 723. (b) Long, E. C.; Barton, J. K. *Acc. Chem. Res.* **1990**, *23*, 2773.
- 9. Binding commonly results in hypochromism and a shift of the transition to longer wavelength in the intercalated chromophore. (a) Dougherty, G.; Pigram, W. J CRC Crit. Rev. Biochem. 1982, 12, 103. (b) Berman, H. M.; Young, P. R. Annu. Rev. Biophys. Bioeng. 1981, 10, 87.
- 10. McGhee, J.D.; von Hippel, P. H. J. Mol. Biol. 1974, 86, 469.
- 11. Cohen, S. N.; Yielding, K. L. J. Biol. Chem. 1965, 240, 3123
- (a) Cory, M.; Tidwell, R. R.; Fairley, T. A. J. Med. Chem. 1992, 35, 431. (b) Fairley, T. A.; Tidwell, R. R.; Donkor, I.; Naimen, N. A.; Ohemeng, K. A.; Lombardy, R. J.; Bentley, J. A.; Cory, M. J. Med. Chem. 1993, 36, 1746.
- 13. All the new products were identified by IR, ¹H NMR, MS and elemental analyses (± 0.4). Data of representative compound **4a**: mp = 283-284 °C (EtOH) IR (KBr): υ_{máx} 1637, 1604, 1404, 1223, 1169, 1083, 1013 cm⁻¹. ¹H-NMR (300 MHz, DMSO-d₆): δ 13.30 (bs, 1H, NH); 9.28 (d, 1H, J=7.3 Hz); 9.22 (s, 1H); 8.88 (d, 1H, J=8.2 Hz); 8.27 (d, 1H, J=7.3 Hz); 7.91 (d, 1H, J=8.2 Hz); 7.74 (t, 1H, J=8.2 Hz, J=7.3 Hz); 7.58 (t, 1H, J=8.2 Hz, J=7.3 Hz); 6.73 (s, 2H); 2.75 (s, 3H); 2.72 (s, 3H); 2.49 (s, 6H); 2.16 (s, 3H) ppm. MS (EI): M/z (relat. int.) 247 (M⁺,100); 205 (26); 182 (14).
- 14. Braña, M. F.; Castellano, J. M.; Morán, M.; Pérez de la Vega, M. J.; Romerdahl, C. A.; Quian, X-D.; Bousquet, P.; Emling, F.; Schlick, E.; Keilhauer, G. Anticancer Drug Res. 1993, 8, 257.
- (a) Insight-II, version 2.2.0, 1994; Biosym Technologies Inc., 9685 Scranton Road, San Diego, CA 92121-2777.
 (b) Gaussian 92, Revision D.2, Frisch, M. J.; Trucks, G. W.; Head-Gordon, M.; Gill, P. M. W.; Wong, M. W.; Foresman, J. B.; Johnson, G.; Schlegel, H. B.; Robb, M. A.; Replogle, E. S.; Gomperts, R.; Andres, J. L.; Raghavachari, K.; Binkley, J. S.; González, C.; Martin, R. L.; Fox, D. J.; Defrees, D.J.; Baker, J.; Stewart, J. J. P.; Pople, J. A.; Gaussian, Inc., Pittsburgh PA, 1992.
- For a detailed discussion of binding parameters for ethidium bromide see: (a) Graves, D. E.; Watkins, C. L.;
 Yielding, L. W. Biochemistry, 1981, 20, 1887. (b) Dougherty, G.; Pigram, J. W. CRC Crit. Rev. Biochem.,
 vol, 12, p. 103-132, 1982.